sequence followed by restriction sites HpaI, ClaI and PmeI and was introduced into BamHI/Asp718 sites. A gene cassette comprising of a "humanized" version of the ORF coding for GFP (gfp-h) under the control of the HCMV enhancer/promoter and the Ptk/PY441 enhancer-driven neoR selection marker was excised from plasmid pUF5 (Zolotukhin et al., 1996, J. Virol. 70, 4646-4654) and inserted into the recombination vector via BglII sites.

A1 cont.

On page 35, delete the paragraph beginning at line 18 and replace it with the following paragraph:

At the 5' and 3'-positions of this loxP-flanked gene cassette, two HCMV sequences with homology to the gene region containing the open reading frames US9 and US10 were inserted. For this, viral sequences were amplified from template pCM49 (Fleckenstein et al., 1982, Gene 18, 39-46) via PCR in a 35-cycle program (denaturation 45 sec at 95°C, annealing 45 sec at 55°C and elongation 2 min at 72°C) by the use of Vent DNA polymerase (New England Biolabs). A US10-specific sequence of 1983 bp in length was generated using primers US10[200900]SpeI (GCTC**ACTAGT**GGCCTAGCCTGGCTCATGGCC) (SEQ ID NO:7) and US10[198918]PacI (GTCC**TTAATTAA**GACGTGGTTGTGGTCACCGAA) (SEQ ID NO:8) and inserted at the vector 5' cloning position via SpeI/PacI restriction sites (see bold-print). A US9-specific sequence of 2010 bp was generated using primers US9-3'PmeI (CTCG**GTTTAAAC**GACGTGAGGCGCTCCGTCACC) (SEQ ID NO:9) and US-5' ClaI (TTGC**ATCGAT**ACGGTGTGAGATACCACGATG) (SEQ ID NO:10) inserted at the vector 3' cloning position via PmeI/ClaI restriction sites.

A2

Please delete the paragraph bridging pages 35 and 36, and replace it with the following paragraph:

The resulting construct pHM673 was linearized by the use of restriction enzyme NheI and transfected into HEF cells via the electroporation method using a Gene Pulser (Bio-rad; 280 V, 960 μF, 400 Ω). After 24 h of cultivation, cells were used for infection with 1 PFU/ml of HCMV strain AD169. Selection with 200 μg/ml G418 was started 24 h post infection. Following 3 weeks of passage in the presence of G418, GFP fluorescence could be detected in most of the infected cells. Plaque assays were performed with infectuous culture supernatant on HFF cells and single virus plaques were grown by transfer to fresh HFF cells cultured in 48-well plates. DNA was isolated from cells of 32 fluorescence-positive wells and confirmed for the presence of recombinant virus by PCR. For this, primers US9[198789] (TGACGCGAGTATTACGTGTC) (SEQ ID NO:11) and US10[199100] (CTCCTCCTGATATGCGGTT) (SEQ ID NO:12) were used resulting in an amplification product of 312 bp for wild-type AD169 virus and approximately 3.5 kb for recombinant virus.

A3